

REVIEW ARTICLE

Clotrimazole as a pharmaceutical: past, present and future.

P.D. Crowley and H.C. Gallagher

School of Medicine and Medical Science, Conway Institute, University College Dublin, Belfield, Dublin, Ireland

Keywords

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CorrespondenceHelen C. Gallagher, School of Medicine and Medical Science, Conway Institute, University College Dublin, Belfield, Dublin 4, Ireland.
E-mail: helen.gallagher@ucd.ie

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Summary

Clotrimazole is a broad-spectrum antimycotic drug mainly used for the treatment of *Candida albicans* and other fungal infections. A synthetic, azole antimycotic, clotrimazole is widely used as a topical treatment for *tinea pedis* (athlete's foot), as well as vulvovaginal and oropharyngeal candidiasis. It displays fungistatic antimycotic activity by targeting the biosynthesis of ergosterol, thereby inhibiting fungal growth. As well as its antimycotic activity, clotrimazole has become a drug of interest against several other diseases such as sickle cell disease, malaria and some cancers. It has also been combined with other molecules, such as the metals, to produce clotrimazole complexes that show improved pharmacological efficacy. Moreover, several new, modified-release pharmaceutical formulations are also undergoing development. Clotrimazole is a very well-tolerated product with few side effects, although there is some drug resistance appearing among immunocompromised patients. Here, we review the pharmaceutical chemistry, application and pharmacology of clotrimazole and discuss future prospects for its further development as a chemotherapeutic agent.

Introduction

Clotrimazole is a broad-spectrum antimycotic drug that is in widespread use for the treatment of *Candida albicans* and other fungal infections. Its antimycotic properties were discovered in the late 1960s. As an active ingredient, it is marketed as a generic drug under various different trade names and by various companies worldwide. In addition to its antimycotic activity, clotrimazole is used in the treatment of metronidazole-resistant *Trichomoniasis* to relieve symptoms (Cudmore *et al.* 2004) and displays activity against certain Gram-positive bacteria (Alsterholm *et al.* 2010). It is a synthetic compound.

Chemical structure and molecular formula

The molecular formula of clotrimazole is $C_{22}H_{17}ClN_2$, and its molecular weight is 344.8 g mol^{-1} . The structure of clotrimazole is illustrated in Fig. 1.

Description of structural features

Clotrimazole is considered to be chemically peculiar (Fig. 1). It contains four aromatic rings bonded to a

tetrahedral (sp^3 hybridized) carbon atom, causing a highly steric encumbrance on this atom. One of the aromatic groups is an imidazole ring, and this is known to mediate electron transfer reactions in biological systems (Eaton and Wilkins 1978; Eaton and Wilson 1979). Its remaining aromatic rings comprise a triphenylmethyl system – a structure that is known to form and stabilize radical intermediates (Hicks 2007). One of these rings is chloro-substituted at its C2 position. Although clotrimazole is an achiral molecule, its two phenyl rings are enantiotopic, with one being pro-R and the other pro-S. These enantiotopic specificities can be differentiated by interaction with a chiral molecule (Eliei *et al.* 1994).

Computational modelling of clotrimazole in a structure-based mechanistic study yielded four stable conformers, none of which has two aromatic rings in the same plane (Navas *et al.* 2004). These computational studies indicated that the energy content of a putative coplanar conformer is very high, resulting in an extremely unstable structure, due to interactions between the substituents at the *ortho*-positions in the aromatic rings. Thus, the authors concluded that clotrimazole does not have the coplanar physical properties that are typical of many xenobiotics that act as ligands for the aryl hydrocarbon receptor, but instead has a

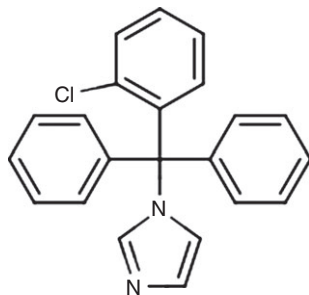


Figure 1 The chemical structure of clotrimazole (1-[(2-chlorophenyl)diphenylmethyl]-1H-imidazole).

‘propeller-like’ conformation. These computed models of clotrimazole’s structure are supported by X-ray diffraction analysis of the crystalline form of clotrimazole (Song and Shin 1998). Navas and co-workers (Navas *et al.* 2004) also computed the molecular electrostatic potential (MEP) and dipole moment of clotrimazole. As these two parameters provide an indication of the charge distribution and electrostatic potential of a molecule, they are used to model and explain the interactions between biologically active chemicals and their biomolecular targets. MEP mapping revealed that clotrimazole possesses a peripheral electron-rich region corresponding to its unsubstituted nitrogen atom and a region with a positive electrostatic potential that corresponds to the substituted nitrogen atom. This analysis suggested that clotrimazole would interact efficiently with acidic or electrophilic species that are present in biological target molecules via its unsubstituted nitrogen. Its dipole moment values were typical of molecules with a high proportion of heteroatoms, low symmetry and relatively large size. For all four conformations of clotrimazole, the dipole orientates from the imidazole ring (negative end) towards the chlorine atom (positive end) and dipole moment values ranged from 3.78 to 5.58 D.

Therapeutic class and pharmaceutical use

Clotrimazole is a member of the azole class of synthetic antimycotic agents that were discovered in the 1960s. Azoles comprise the largest class of antimycotic drugs in clinical use and can be further subdivided into two classes on the basis of their chemical structure: imidazoles and triazoles. Clotrimazole falls into the imidazole subclass of azole drugs. Along with econazole and miconazole, clotrimazole is the drug of choice for the topical treatment of tinea pedis (athlete’s foot), tinea cruris and tinea corporis caused by isolates of *Trichophyton rubrum*, *Trichophyton mentagrophytes*, *Epidermophyton floccosum*, *Microsporum canis* and *C. albicans* (Gelone and O’Donnell 2006). It is also widely used in the topical treatment of vulvovaginal and oropharyngeal candidiasis.

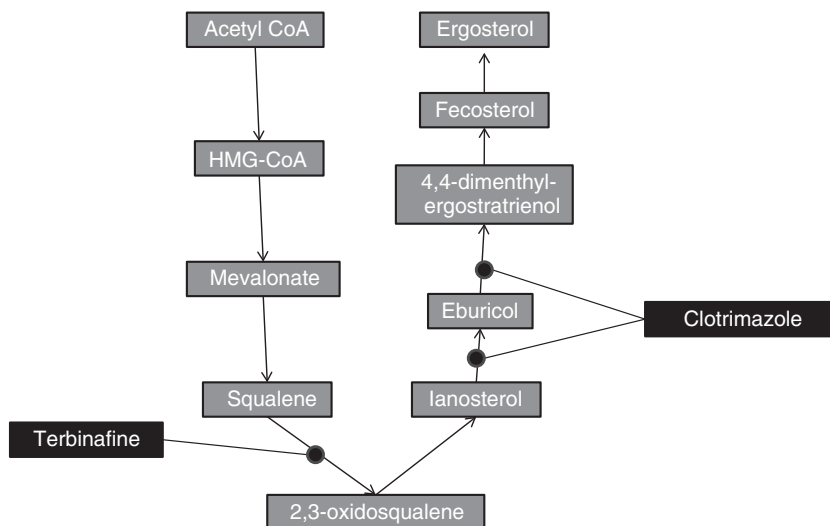
Brief summary of antimicrobial activity

All azole-type antimycotic drugs interfere with the biosynthesis of ergosterol, which is a major component of the fungal cytoplasmic membrane. Specifically, azoles including clotrimazole inhibit the microsomal cytochrome P450 (CYP450)-dependent event 14- α -lanosterol demethylation, which is a vital step in ergosterol biosynthesis by fungi (Hitchcock *et al.* 1990) (Fig. 2). The resultant depletion of ergosterol and its replacement with the aberrant sterol species, 14- α -methylsterol, perturb normal membrane permeability and fluidity. Downstream effects include decreased activity of membrane-bound enzymes, including those involved in cell wall synthesis, increased cell wall leakiness and leaking of cell contents. Moreover, because ergosterol directly stimulates growth of fungal cells in a hormone-like fashion, rapid onset of these events results in a dose- and time-dependent inhibition of fungal growth. Clotrimazole is generally considered to be a fungistatic rather than a fungicidal drug, although as for many antimicrobials, this distinction is not absolute as it exhibits fungicidal effects at higher concentrations.

The selectivity of azole drugs for fungal cells *in vivo* reflects their greater affinity for fungal versus mammalian cytochrome P450 enzymes. Specifically, these agents target the cytochrome P450-Erg11p or Cyp51p enzyme, which possesses monooxygenase activity and catalyses the removal of the 14- α -methyl group of lanosterol and/or eburicol in fungi. To date, only one Cyp51p crystal structure has been reported – that isolated from *Mycobacterium tuberculosis* (Podust *et al.* 2001). However, by examining this crystal structure, it was possible to identify substrate recognition sites on this protein that could account for the activity of azole antimycotics. Cyp51p proteins contain an iron protoporphyrin moiety located at their active site, and it is at this site that azole compounds bind to the iron atom, via a nitrogen atom (N-3 or N-4) in their imidazole (N-3) or triazole (N-4) ring. The other part of the azole drug binds to an apoprotein, and this interaction is dependent on the individual structure of the drug itself. The exact conformation of this active site differs widely between fungal species and among the many mammalian P450 enzymes (Marichal *et al.* 1990), and, as described below, exploiting these differences offers the possibility of rationally designing drugs with improved efficacy against certain fungal species.

Clotrimazole resistance is a problem particularly in immunocompromised patient populations (Pelletier *et al.* 2000; Sobel 2007). Resistance has been linked to overexpression of efflux pump genes such as the *Candida* drug resistance₁, *Candida* drug resistance₂ and multidrug resistance₁ genes (White *et al.* 2002). In *Candida glabrata*, the protein antiporter, CgTpo3, was found to increase

Figure 2 Ergosterol is a vital component of fungal plasma membranes fulfilling a similar function to cholesterol in animal cell membranes. Clotrimazole targets the enzyme lanosterol 14- α -demethylase responsible for the conversion of lanosterol to 4,4-dimethyl-ergostrienol a common target of azole drugs. Other antifungal drugs, such as terbinafine, target squalene epoxidase.



resistance to azole drugs (Costa *et al.* 2014). Changes in the drug target lanosterol 14- α -demethylase caused by mutations to, or overexpression of, the ERG11 gene may also cause resistance in some cases (Mishra *et al.* 2007). In some cases of resistance, nonazole treatments or second-generation drugs may be suitable as alternative pharmacotherapies (Valerio *et al.* 2013).

While CYP450 inhibition is accepted to primarily account for the antimycotic properties of clotrimazole, this drug also exhibits other diverse pharmacological actions. These include inhibition of sarcoplasmic reticulum Ca²⁺-ATPase (Bartolommei *et al.* 2006), depletion of intracellular calcium stores (Jan *et al.* 2000) and blockade of calcium-dependent potassium channels and voltage-dependent calcium channels (Rittenhouse *et al.* 1997; Wu *et al.* 1999; Shah *et al.* 2001; Tian *et al.* 2006). It is believed that the action of clotrimazole on these diverse targets accounts for biological effects of this drug that are independent of its antimycotic action. For example, clotrimazole inhibits the proliferation of several normal and cancer cell lines *in vitro* (Benzaquen *et al.* 1995) and inhibits the expression of adhesion molecules by TNF- α (Thapa *et al.* 2009), and it has been reported to exert neuroprotective effects and to modify the cytotoxicity of some metal cations (Oyama *et al.* 2006). There is much interest in the therapeutic application of clotrimazole in sickle cell disease, because its metabolite, ICA 17043, is believed to exert a beneficial effect on erythrocyte dehydration by blocking the Gardos calcium channel (Brugnara *et al.* 1996; Brugnara and De Franceschi 2006; Gbotosho *et al.* 2013). Clotrimazole also exhibits antimalarial activity *in vitro* (Tiffert *et al.* 2000), possibly by inhibiting hemoperoxidase, thereby causing oxidative stress in the parasite (Trivedi *et al.* 2005).

Similar drugs

Antimycotic azole drugs that are closely related to clotrimazole include the imidazole drugs, miconazole, econazole and ketoconazole, and the triazole drugs fluconazole and itraconazole. They are all broad-spectrum antimycotics and are generally poorly absorbed orally, with the exception of ketoconazole and itraconazole (reviewed in Odds *et al.* 2003; Gelone and O'Donnell 2006; Lorand and Kocsis 2007). In the clinical setting, miconazole and econazole are used very similarly to fluconazole in the treatment of topical fungal infections, whereas miconazole is also frequently used intravenously for the treatment of systemic infections in patients who cannot tolerate amphotericin. Unlike other azoles, itraconazole displays activity against *Aspergillus*.

Because the extent of antifungal activity for each azole drug depends largely on the exact interaction between the drug and cytochrome P450, rational drug design approaches have been employed in the design of a new generation of azole drugs that incorporate triazole structures (reviewed in Odds *et al.* 2003; Lorand and Kocsis 2007; Weig and Brown 2007; Pasqualotto *et al.* 2010). These newer drugs include voriconazole and ravuconazole, which are structurally based on fluconazole and posaconazole, which is closely related to itraconazole. In comparison with the first-generation triazole drugs, the newer triazole compounds have extended antimycotic spectrums. These drugs have, in essence, been developed to address the limitations of existing azole compounds, including the emergence of drug-resistant fungal strains. Use of these second-generation drugs is reviewed in the context of paediatric medicine in Valerio *et al.* (2013).

Two other classes of antimycotic drugs also target the ergosterol biosynthetic pathway, despite being considered chemically distinct from clotrimazole or azole compounds in general. Most notably, the allylamines, including terbinafine, inhibit the enzyme squalene epoxidase which acts earlier in the ergosterol biosynthetic pathway (Fig. 2). These compounds can have fungicidal effects and have also demonstrated activity against filamentous fungi and a few pathogenic yeasts. Thus, there is considerable interest in the possibility of formulating them in combination with azole drugs to achieve synergistic inhibitory effects on ergosterol synthesis. The phenylmorpholine class of drugs that includes amoroline affects two late events in ergosterol synthesis by inhibiting Erg24p reductase and Erg2p isomerase enzymes. It is largely used for superficial mycoses, and its targets have not been researched extensively.

Pharmaceutical dosage forms and administration

Within the European Union, clotrimazole is available in topical cream and pessary formulations under a variety of trade names. In the USA, additional formulations are available, including clotrimazole lotions, powders, lozenges, topical solutions and vaginal inserts/tablets. Clotrimazole is sometimes formulated with the steroids hydrocortisone or bethamethasone, and in some cases, these compounded preparations may be labelled as coclimasone (Sweetman 2007). Typical excipients in clotrimazole creams include benzyl alcohol, cetostearyl alcohol, medium-chain triglycerides and tricetareth-4 phosphate.

Although its availability differs slightly from country to country, monoprparations of clotrimazole are generally available over-the-counter, while combined preparations may require a prescription. In products aimed at the treatment of fungal skin infections, clotrimazole is usually formulated as a 1% cream, lotion, spray or solution. In the treatment of vulvovaginal candidiasis, the normal dosage forms are either 100 mg, 200 mg or 500 mg pessaries which are administered daily for 6, 3 or 1 days, respectively. Similar doses may be obtained by application of 1, 2 or 10% creams to the vaginal area. In the treatment of oropharyngeal candidiasis, clotrimazole lozenges are usually formulated to contain 10 mg of the active drug and these are sucked slowly until dissolved, five times daily for 14 days. In the prophylactic prevention of oropharyngeal candidiasis in immunosuppressed individuals, this dose is reduced to 10 mg, three times daily, for the duration of the immunosuppressive therapy.

According to the USP, clotrimazole should contain not less than 98.0% and not more than 102.0% clotrimazole while clotrimazole creams, lotions, lozenges, topical solutions and vaginal inserts should contain not less than 90.0% and not more than 110.10% of the labelled

amount of clotrimazole. According to the British and European Pharmacopoeias, clotrimazole should contain not less than 98.5% and not more than 100.5% of clotrimazole with reference to the dried substance.

Side effects, interactions and contraindications

Topical forms of clotrimazole are available as over-the-counter medication and are considered reasonably safe and without serious side effects. However, there have been limited case reports of contact allergic dermatitis with clotrimazole creams that are not attributable to allergies to the vehicles or excipients, but are caused by the active ingredient itself (Kalb and Grossman 1985). Intravaginal clotrimazole applied via a pessary may damage latex contraceptives (condoms) necessitating the use of additional contraceptive measures during the period of administration.

The most prominent side effects of clotrimazole preparations in current use are those associated with the use of oral lozenges for the treatment of oral candidiasis. These include nausea, vomiting, unpleasant mouth sensations, pruritus and elevation of liver enzymes (reviewed in Ellepola and Samaranyake 2000). Clotrimazole lozenges are not sold in the European Union but are widely available in the USA and other countries. Clotrimazole tablets or capsules designed for swallowing, as opposed to sucking, are no longer used as they were associated with GIT disturbances, dysuria and mental depression. Notably, because of clotrimazole's very limited water solubility and GIT toxicity, other imidazole antifungal drugs have replaced clotrimazole in oral capsule formulations. For example, oral Canesten, which is available over-the-counter in the United Kingdom, but not in Ireland, contains ketoconazole.

Because clotrimazole is not systemically absorbed, drug interactions are not a major issue with its use. It can be used safely with consumption of alcohol, does not affect driving ability, and there is no evidence of it posing a risk to the developing foetus in pregnancy. Pessaries are not recommended for use in children or infants, although the drug itself poses no special risk to this subpopulation. Clotrimazole is also safe for use in the elderly population and in breast-feeding mothers.

Concerns surrounding environmental toxicity

There are growing concerns about the abundance of chemicals in our environment for which inadequate data are available with regard to their toxicity. While pharmaceutical compounds are stringently assessed with regard to their toxicity to humans prior to their marketplace release, there is relatively little assessment of their environmental effects. Certain types of chemicals are considered to be persistent in the environment as they can

bioaccumulate in wildlife species. Aquatic wildlife is at particular risk as it is exposed to discharges from municipal waste water from households and industry. Chemicals that affect key enzymatic pathways that are evolutionarily conserved across species, such as cytochrome P450, are of particular concern as they may have unintentional ecotoxicological effects. Clotrimazole fits several of these criteria as it targets cytochrome P450 activity, is nonbiodegradable in the environment (i.e. half-life of more than 60 days) and is therefore considered to be a persistent chemical. As a consequence, clotrimazole was included on the OSPAR list of chemicals for priority action at the meeting of the Convention for the Protection of the Marine Environment of the north-east Atlantic (the OSPAR Convention) in 2002. However, after conducting a thorough environmental risk assessment of clotrimazole, it was concluded that clotrimazole does not pose a significant environmental risk to marine and aquatic life (OSPAR Commission, 2005). Despite this, some recent studies have suggested that clotrimazole may affect marine microalgae at low concentrations (Porsbring *et al.* 2009).

Conclusion and future directions

Invasive fungal infections have increased in frequency worldwide in recent decades and have emerged as a major cause of illness and death, especially in immunocompromised individuals (Malani and Kauffman 2007). One reason for the marked increase in these problematic infections is the growing size of the highly immunocompromised patient population. Ironically, this trend in part reflects clinical success in other areas such as treatment of human immunodeficiency virus (HIV) infection, cancer and increasing numbers of transplant recipients. Resistance to clotrimazole, which used to be a rare occurrence, is now quite common in certain patient subpopulations with candidiasis (Pelletier *et al.* 2000). While these epidemiological trends are unlikely to impact significantly on the widespread use of clotrimazole and related drugs in the general population, they do continue to drive the rational design of new drug entities with improved activity spectrums and these newer drugs, such as posaconazole, are likely to supersede clotrimazole for the treatment of invasive fungal infections in select, high-risk patient populations.

Nevertheless, the further development of clotrimazole as a pharmaceutical is an area of intense research at present. There are prospects both for its exploitation in new indications and for the development of new formulations. A scaffold based on clotrimazole is being used as a pharmacophore in the design and synthesis of novel antimalarial drugs that are cheap and easy to synthesize (Gemma *et al.* 2007, 2008). Palladium–clotrimazole complexes that exhibit enhanced cytotoxicity against tumour

cell lines, in comparison with clotrimazole alone, are under investigation as novel antineoplastic agents (Navarro *et al.* 2006). Indeed, several other metal–clotrimazole complexes, such as ruthenium–clotrimazole and platinum–clotrimazole, also display promising antineoplastic characteristics (Navarro *et al.* 2009; Robles-Escajeda *et al.* 2013). There is also intense interest in using clotrimazole and its metabolite as lead compounds in the strategic design of novel treatments for sickle cell disease, on the basis that they can reduce erythrocyte dehydration *in vivo* by inhibiting the so-called Gardos, calcium-dependent potassium channel that malfunctions in this disease (Brugnara *et al.* 1996; Brugnara and De Franceschi 2006; Gbotosho *et al.* 2013).

New approaches to formulation of clotrimazole include a buccal bioadhesive film containing clotrimazole, which was found to inhibit oral candidiasis for up to 6 h (Singh *et al.* 2008), and a thermosensitive vaginal gel formulation formed by complexation of clotrimazole with beta-cyclodextrin, which has been shown to reduce the release rate of clotrimazole in comparison with standard preparations (Bilensoy *et al.* 2006). This type of slow-release formulation may exhibit increased efficacy over other vaginal delivery systems, as traditional vaginal creams, pessaries and tablets tend to have short residency times in the vagina due to the natural cleansing process that takes place there. The use of liposomes containing clotrimazole may also provide increased residency in the vagina, thereby improving gel formulations for treatment (Vanić and Škalko-Basnet 2013). RS 100 nano-capsules have recently been studied in the treatment of *C. albicans* and *C. glabrata*, and these have been reported as more active than free clotrimazole alone (Santos *et al.* 2014). Given the scale of the current market for vulvovaginal clotrimazole preparations, novel formulations that can demonstrate advantage over pre-existing preparations could potentially attract a large revenue stream. Nano-fibre mats for oral applications are also superior in efficacy and have reduced toxicity over lozenges and powders in current use, although further pharmaceuticals investigations are needed (Tonglairoum *et al.* 2014).

In conclusion, clotrimazole is an effective, safe and well-tolerated drug with an unusual chemistry that is widely used in the treatment of skin, vulvovaginal and oropharyngeal fungal infections. It is sold in most developed countries worldwide under a variety of trade names, and a large number of clotrimazole formulations are available. Although emerging resistance to clotrimazole may limit the future use of this drug in certain patient subpopulations, in the general population, its widespread use is likely to continue for the foreseeable future. Ongoing development of clotrimazole as a pharmaceutical is currently focused on finding new clinical indications for the drug, its use as a lead compound in structure-based drug

design studies and the optimization of formulated products to enhance drug delivery.

Conflict of Interest

No conflict of interest declared.

References

- Alsterholm, M., Karami, N. and Faergemann, J. (2010) Antimicrobial activity of topical skin pharmaceuticals – an in vitro study. *Acta Derm Venereol* **90**, 239–245.
- Bartolommei, G., Tadini-Buoninsegni, F., Hua, S., Moncelli, M.R., Inesi, G. and Guidelli, R. (2006) Clotrimazole inhibits the Ca²⁺-ATPase (SERCA) by interfering with Ca²⁺ binding and favoring the E2 conformation. *J Biol Chem* **281**, 9547–9551.
- Benzaquen, L.R., Brugnara, C., Byers, H.R., Gattoni-Celli, S. and Halperin, J.A. (1995) Clotrimazole inhibits cell proliferation in vitro and in vivo. *Nat Med* **1**, 534–540.
- Bilensoy, E., Rouf, M., Vural, I. and Hincal, A. (2006) Thermosensitive vaginal gel formulation for the controlled release of clotrimazole via complexation to beta-cyclodextrin. *J Control Release* **116**, e107–e109.
- Brugnara, C. and De Franceschi, L. (2006) Clinical trials of new therapeutic pharmacology for sickle cell disease. *Sante* **16**, 263–268.
- Brugnara, C., Gee, B., Armsby, C.C., Kurth, S., Sakamoto, M., Rifai, N., Alper, S.L. and Platt, O.S. (1996) Therapy with oral clotrimazole induces inhibition of the Gardos channel and reduction of erythrocyte dehydration in patients with sickle cell disease. *J Clin Invest* **97**, 1227–1234.
- Costa, C., Nunes, J., Henriques, A., Mira, N.P., Nakayama, H., Chibana, H. and Teixeira, M.C. (2014) *Candida glabrata* drug: H⁺ antiporter CgTpo3 (ORF CAGL0I10384g): role in azole drug resistance and polyamine homeostasis. *J Antimicrob Chemother.* doi:10.1093/jac/dku1044. [Epub ahead of print].
- Cudmore, S.L., Delgaty, K.L., Hayward-McClelland, S.F., Petrin, D.P. and Garber, G.E. (2004) Treatment of infections caused by metronidazole-resistant *Trichomonas vaginalis*. *Clin Microbiol Rev* **17**, 783–793.
- Eaton, D.R. and Wilkins, R.G. (1978) Reduction by dithionite of adducts of metmyoglobin with imidazole, pyridine, and derivatives. *J Biol Chem* **253**, 908–915.
- Eaton, D. and Wilson, K. (1979) Reaction of imidazole and hydroquinone with oxymyoglobin. *J Inorg Biochem* **10**, 195–203.
- Eliel, E.L., Wilen, S.H. and Mander, L.N.(eds) (1994) . *Stereochemistry of organic compounds*. p. 381. New York: John Wiley and Sons.
- Ellepol, A. and Samaranyake, L. (2000) Oral candidal infections and antimycotics. *Crit Rev Oral Biol Med* **11**, 172–198.
- Gbotosho, O.T., Cytlak, U.M., Hannemann, A., Rees, D.C., Tewari, S. and Gibson, J.S. (2013) Inhibitors of second messenger pathways and Ca-induced exposure of phosphatidylserine in red blood cells of patients with sickle cell disease. *Pflugers Arch.* doi:10.1007/s00424-00013-01343-00428. [Epub ahead of print].
- Gelone, S.A. and O'Donnell, J. (2006) Anti infectives. In *Remington the science and practice of pharmacy*, 21st Edition ed. Troy, D.B., pp. 1626–1684. Baltimore: Lippincott Williams and Wilkins.
- Gemma, S., Campiani, G., Butini, S., Kukreja, G., Joshi, B.P., Persico, M., Catalanotti, B., Novellino, E. et al. (2007) Design and synthesis of potent antimalarial agents based on clotrimazole scaffold: exploring an innovative pharmacophore. *J Med Chem* **50**, 595–598.
- Gemma, S., Campiani, G., Butini, S., Kukreja, G., Coccone, S.S., Joshi, B.P., Persico, M., Nacci, V. et al. (2008) Clotrimazole scaffold as an innovative pharmacophore towards potent antimalarial agents: design, synthesis, and biological and structure–activity relationship studies. *J Med Chem* **51**, 1278–1294.
- Hicks, R.G. (2007) What's new in stable radical chemistry? *Org Biomol Chem* **5**, 1321–1338.
- Hitchcock, C.A., Dickinson, K., Brown, S., Evans, E. and Adams, D. (1990) Interaction of azole antifungal antibiotics with cytochrome P-450-dependent 14 alpha-sterol demethylase purified from *Candida albicans*. *Biochem J* **266**, 475–480.
- Jan, C.-R., Tseng, C.-J., Chou, K.-J. and Chiang, H.-T. (2000) Novel effects of clotrimazole on Ca²⁺ signaling in Madin Darby canine kidney cells. *Life Sci* **66**, 2289–2296.
- Kalb, R. and Grossman, M. (1985) Contact dermatitis to clotrimazole. *Cutis* **36**, 240–242.
- Lorand, T. and Kocsis, B. (2007) Recent advances in antifungal agents. *Mini-Rev Med Chem* **7**, 900–911.
- Malani, A.N. and Kauffman, C.A. (2007) Changing epidemiology of rare mould infections. *Drugs* **67**, 1803–1812.
- Marichal, P., Gorrens, J. and Coene, M. (1990) Biochemical basis for the activity and selectivity of oral antifungal drugs. *Br J Clin Pract Suppl* **71**, 41–46.
- Mishra, N., Prasad, T., Sharma, N., Payasi, A., Prasad, R., Gupta, D. and Singh, R. (2007) Pathogenicity and drug resistance in *Candida albicans* and other yeast species. *Acta Microbiol Immunol Hung* **54**, 201–235.
- Navarro, M., Peña, N.P., Colmenares, I., González, T., Arsenak, M. and Taylor, P. (2006) Synthesis and characterization of new palladium–clotrimazole and palladium–chloroquine complexes showing cytotoxicity for tumor cell lines in vitro. *J Inorg Biochem* **100**, 152–157.
- Navarro, M., Higuera-Padilla, A.R., Arsenak, M. and Taylor, P. (2009) Synthesis, characterization, DNA interaction studies and anticancer activity of platinum–clotrimazole complexes. *Transition Met Chem* **34**, 869–875.
- Navas, J.M., Chana, A., Herradón, B. and Segner, H. (2004) Induction of cytochrome P4501A (CYP1A) by clotrimazole, a non-planar aromatic compound. Computational studies

- on structural features of clotrimazole and related imidazole derivatives. *Life Sci* **76**, 699–714.
- Odds, F.C., Brown, A.J. and Gow, N.A. (2003) Antifungal agents: mechanisms of action. *Trends Microbiol* **11**, 272–279.
- OSPAR Commission (2005) Hazardous Substance Series: Open background documentation on clotrimazole; publication no 2005/199. Available at: http://www.ospar.org/documents/dbase/publications/p00199/p00199_bd%20on%20clotrimazole.pdf. (accessed 10 June 2014).
- Oyama, T.M., Oyama, T.B., Oyama, K., Matsui, H., Horimoto, K., Nishimura, Y. and Oyama, Y. (2006) Clotrimazole, an antifungal drug possessing diverse actions, increases the vulnerability to cadmium in lymphocytes dissociated from rat thymus. *Toxicology* **228**, 269–279.
- Pasqualotto, A.C., Thiele, K.O. and Goldani, L.Z. (2010) Novel triazole antifungal drugs: focus on isavuconazole, ravuconazole and albaconazole. *Curr Opin Investig Drugs*, **11**, 165–174.
- Pelletier, R., Peter, J., Antin, C., Gonzalez, C., Wood, L. and Walsh, T.J. (2000) Emergence of resistance of *Candida albicans* to clotrimazole in human immunodeficiency virus-infected children: in vitro and clinical correlations. *J Clin Microbiol* **38**, 1563–1568.
- Podust, L.M., Poulos, T.L. and Waterman, M.R. (2001) Crystal structure of cytochrome P450 14 α -sterol demethylase (CYP51) from *Mycobacterium tuberculosis* in complex with azole inhibitors. *Proc Natl Acad Sci* **98**, 3068–3073.
- Porsbring, T., Blanck, H., Tjellström, H. and Backhaus, T. (2009) The pharmaceutical clotrimazole affects marine microalgal communities at picomolar concentrations. In *SETAC Europe 19th Annual Meeting, Gothenburg, Sweden*. <http://swepubkbse/bib/swepub:oai:service.scigoorg:113304?tab=2&abs&language=en>.
- Rittenhouse, A., Vandorpe, D., Brugnara, C. and Alper, S. (1997) The antifungal imidazole clotrimazole and its major in vivo metabolite are potent blockers of the calcium-activated potassium channel in murine erythroleukemia cells. *J Membr Biol* **157**, 177–191.
- Robles-Escajeda, E., Martínez, A., Varela-Ramirez, A., Sánchez-Delgado, R.A. and Aguilera, R.J. (2013) Analysis of the cytotoxic effects of ruthenium–ketoconazole and ruthenium–clotrimazole complexes on cancer cells. *Cell Biol Toxicol* **29**, 431–443.
- Santos, S.S., Lorenzoni, A., Pegoraro, N.S., Denardi, L.B., Alves, S.H., Schaffazick, S.R. and Cruz, L. (2014) Formulation and in vitro evaluation of coconut oil-core cationic nanocapsules intended for vaginal delivery of clotrimazole. *Colloids Surf B Biointerfaces* **116c**, 270–276.
- Shah, M., Miscony, Z., Javadzadeh-Tabatabaie, M., Ganellin, C. and Haylett, D. (2001) Clotrimazole analogues: effective blockers of the slow afterhyperpolarization in cultured rat hippocampal pyramidal neurones. *Br J Pharmacol* **132**, 889–898.
- Singh, S., Jain, S., Muthu, M., Tiwari, S. and Tilak, R. (2008) Preparation and evaluation of buccal bioadhesive films containing clotrimazole. *AAPS PharmSciTech* **9**, 660–667.
- Sobel, J.D. (2007) Vulvovaginal candidosis. *Lancet* **369**, 1961–1971.
- Song, H. and Shin, H.-S. (1998) The antifungal drug clotrimazole. *Acta Crystallogr Sect C: Cryst Struct Commun* **54**, 1675–1677.
- Sweetman, S.C. (eds) (2007) . *Martindale: the complete drug reference*. p. 764. London: Pharmaceutical Press.
- Thapa, D., Lee, J.S., Park, M.-A., Cho, M.-Y., Park, Y.-J., Choi, H.G., Jeong, T.C. and Kim, J.-A. (2009) Inhibitory effects of clotrimazole on TNF- α -induced adhesion molecule expression and angiogenesis. *Arch Pharm Res* **32**, 593–603.
- Tian, M., Dong, M.Q., Chiu, S.W., Lau, C.P. and Li, G.R. (2006) Effects of the antifungal antibiotic clotrimazole on human cardiac repolarization potassium currents. *Br J Pharmacol* **147**, 289–297.
- Tiffert, T., Ginsburg, H., Krugliak, M., Elford, B.C. and Lew, V.L. (2000) Potent antimalarial activity of clotrimazole in in vitro cultures of *Plasmodium falciparum*. *Proc Natl Acad Sci* **97**, 331–336.
- Tonglairoum, P., Ngawhirunpat, T., Rojanarata, T., Kaomongkolgit, R. and Opanasopit, P. (2014) Fast-acting clotrimazole composited PVP/HPbetaCD nanofibers for oral candidiasis application. *Pharm Res*. doi:10.1007/s11095-11013-11291-11091. [Epub ahead of print].
- Trivedi, V., Chand, P., Srivastava, K., Puri, S.K., Maulik, P.R. and Bandyopadhyay, U. (2005) Clotrimazole inhibits hemoperoxidase of *Plasmodium falciparum* and induces oxidative stress. Proposed antimalarial mechanism of clotrimazole. *J Biol Chem* **280**, 41129–41136.
- Valerio, C., Perillo, T., Brescia, L. and Russo, F. (2013) Antifungal agents in current pediatric practice. *Curr Infect Dis Rep* **15**, 278–287.
- Vanić, Ž. and Škalko-Basnet, N. (2013) Nanopharmaceuticals for improved topical vaginal therapy: can they deliver? *Eur J Pharm Sci* **50**, 29–41.
- Weig, M. and Brown, A.J. (2007) Genomics and the development of new diagnostics and anti-*Candida* drugs. *Trends Microbiol* **15**, 310–317.
- White, T.C., Holleman, S., Dy, F., Mirels, L.F. and Stevens, D.A. (2002) Resistance mechanisms in clinical isolates of *Candida albicans*. *Antimicrob Agents Chemother* **46**, 1704–1713.
- Wu, S.N., Li, H.F., Jan, C.R. and Shen, A.Y. (1999) Inhibition of Ca²⁺ activated K⁺ current by clotrimazole in rat anterior pituitary GH³ cells. *Neuropharmacology* **38**, 979–989.

Other resources

Chemical Drawing package used: MarvinSketch ver. 4.1.13. for Macintosh Downloadable from www.chemaxon.com.